



# The rational design of a novel potent analogue of the 5'-AMP-activated protein kinase inhibitor compound C with improved selectivity and cellular activity

Fouzia Machrouhi<sup>a</sup>, Nouara Ouhamou<sup>a</sup>, Keith Laderoute<sup>b</sup>, Joy Calaoagan<sup>b</sup>, Marina Bukhtiyarova<sup>a</sup>, Paula J. Ehrlich<sup>a</sup>, Anthony E. Klon<sup>a,\*</sup>

<sup>a</sup> Ansaris, Four Valley Square, 512 East Township Line Rd, Blue Bell, PA 19422, United States

<sup>b</sup> SRI International, Menlo Park, CA 94025, United States

## ARTICLE INFO

### Article history:

Received 10 August 2010

Revised 14 September 2010

Accepted 15 September 2010

Available online 19 September 2010

### Keywords:

Fragment-based drug design

FBDD

5'-AMP-activated protein kinase

AMPK

Compound C

## ABSTRACT

We have designed and synthesized analogues of compound C, a non-specific inhibitor of 5'-AMP-activated protein kinase (AMPK), using a computational fragment-based drug design (FBDD) approach. Synthesizing only twenty-seven analogues yielded a compound that was equipotent to compound C in the inhibition of the human AMPK (hAMPK)  $\alpha 2$  subunit in the heterotrimeric complex in vitro, exhibited significantly improved selectivity against a subset of relevant kinases, and demonstrated enhanced cellular inhibition of AMPK.

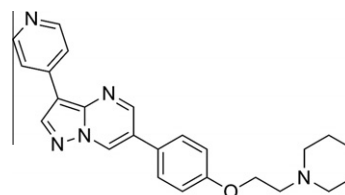
© 2010 Elsevier Ltd. All rights reserved.

5'-AMP-activated protein kinase (AMPK) is a major regulator of normal cellular energy metabolism.<sup>1</sup> AMPK activity is sensitive to the cellular AMP:ATP ratio, which is increased by physiological stresses such as decreased oxygen and glucose concentrations (hypoxia and hypoglycemia). AMPK activity is also associated with the growth of solid tumors, which commonly contain regions of pathologic hypoxia and hypoglycemia.<sup>2</sup> AMPK cooperates with hypoxia-inducible factor-1 (HIF-1), the primary transcriptional regulator of the mammalian response to hypoxia and other metabolic stresses that lower cellular ATP levels, in pathophysiological tumor microenvironments.<sup>3,4</sup> Thus, in the context of microenvironmental stress, AMPK may represent a novel target for the treatment of solid tumors. The successful design of potent and selective inhibitors of AMPK that exhibit activity is an important first step toward establishing the proof-of-concept for the therapeutic value of AMPK inhibition in solid tumor microenvironments.

This Letter describes our efforts to design a Type I inhibitor of AMPK with a superior biological profile than that of compound C (Fig. 1), which is a commonly used experimental direct inhibitor of the enzyme,<sup>5</sup> through the use of our proprietary fragment-based drug design (FBDD) software<sup>6–9</sup> and Imagirotm,<sup>10</sup> our proprietary torsion-space molecular mechanics software package. Compound

C ([4-(2-piperidin-1-yl-ethoxy)-phenyl]-3-pyridin-4-yl-pyrazolo[1,5-a]-pyrimidine) is considered to act as a competitive inhibitor of ATP binding to the catalytic  $\alpha$  subunit of AMPK.<sup>5,11</sup> However, compound C has significantly limited specificity as an AMPK inhibitor.<sup>5</sup>

We discuss the SAR of a series of pyrazolopyrimidine and aminooxazole analogues of compound C and present their general binding mode obtained through the use of homology modeling and Grand Canonical Monte Carlo (GCMC) fragment simulations.<sup>9</sup> A focused kinase selectivity profile for the most active subset of these compounds is presented, as well as the cellular AMPK inhibition data for the most promising pyrazolopyrimidine.



Compound C  
hAMPK IC<sub>50</sub> (μM) = 0.04

Figure 1. Chemical structure of compound C.

\* Corresponding author. Tel.: +1 215 358 2029; fax: +1 215 358 2020.

E-mail address: [aklon@ansarisbio.com](mailto:aklon@ansarisbio.com) (A.E. Klon).

We generated a model of the catalytic  $\alpha 2$  subunit of AMPK with the DFG-loop in the 'in' conformation using MOE.<sup>12</sup> The crystal structure of the AMPK  $\alpha 2$  subunit (RCSB ID# 2H6D, 1.85 Å)<sup>13</sup> was used as the overall template for the protein model. The activation loop (A-loop) was not crystallographically resolved in this structure and the DFG loop is in the 'out' conformation. Residues 157–179 of the microtubule affinity regulating kinase-1 (MARK-1) crystal structure (RCSB ID# 2HAK, 2.60 Å)<sup>14</sup> were used as a template for the 'in' conformation of the DFG-loop and the missing residues for the A-loop. MARK-1 was selected as the template for the A-loop conformation of the AMPK  $\alpha 2$  subunit due to the 50% sequence identity between the AMPK and MARK-1 kinase domains. Compound C, which inhibits AMPK with reported IC<sub>50</sub> values ranging from 0.1 to 0.2  $\mu\text{M}$ <sup>15</sup> was used to model the inhibitor-bound protein complex. The three fragments comprising the core structure of compound C (benzene, pyridine, and pyrazolo[1,5-*a*]pyrimidine, Fig. 1) were simulated for binding to the AMPK homology model using our Grand Canonical Monte Carlo (GCMC) simulation approach.<sup>6–8</sup> Compound C was assembled from the resulting fragment simulation data and found to bind in the ATP site. The resulting complex was subjected to energy minimization, which was terminated when the energy gradient reached 0.001, followed by 259 ps of molecular dynamics simulations in torsion space using Imagi<sup>10</sup> to ensure that the kinetic and potential energies of the system were equilibrated, arriving at the optimized homology model (Fig. 2A).

In the final model, the pyrazolo[1,5-*a*]pyrimidine core was observed to present one ring nitrogen to act as a hydrogen bond

acceptor with the backbone nitrogen of residue Val-96 in the hinge region, while the 4-pyridine fragment forms a hydrogen bond with the side chain of Lys-45. The piperidine fragment is oriented towards the solvent and does not appear to form specific interactions with AMPK. We hypothesize that this fragment performs the function of a solubilizing group. The P-loop (residues 23–30) is moved slightly towards the N-terminal lobe of the kinase in the final model relative to the 2H6D crystal structure of AMPK to allow for better steric contact with the pyridinyl moiety of compound C. The overall RMSD of the C $\alpha$  atoms between the two structures is 2.4 Å.

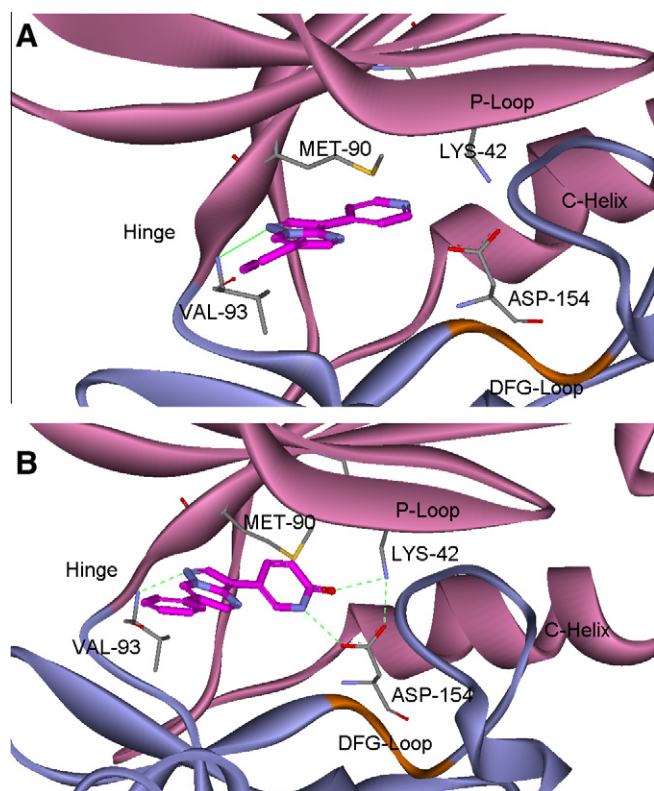
In order to design inhibitors of AMPK, the binding of 1785 small molecular fragments<sup>16</sup> was simulated against the homology model using the GCMC approach. Hypothetical compounds were designed using the results of the fragment simulations to find replacements for the pyridine fragment that maximized interactions with Lys-45 as well as Asp-157 in the DFG-loop as shown in Figure 2B. Replacements for the pyrazolo[1,5-*a*]pyrimidine fragment were explored, with the most successful being aminooxazole. All designed compounds were evaluated using computational ADME filters<sup>17</sup> and prioritized for synthesis on the basis of their calculated binding affinities and ADME properties.

The synthesis of designed substituted pyrazolo[1,5-*a*]pyrimidine derivatives was accomplished according to Scheme 1. The bromide intermediate **2** was readily obtained from the commercially available **1**<sup>18</sup> via an SN2 type reaction with 4-(2-chloroethyl) morpholine hydrochloride salt under basic conditions. The bromide **2** was then reacted with selected commercial available or readily synthesized boronates or boronic acids in microwave palladium catalyzed Suzuki cross-coupling reactions with good to moderate yields to give compounds **3–10**<sup>19</sup> Compound **11** was obtained after a subsequent Suzuki coupling reaction between intermediate **15** and bromide **2** followed by deprotection of the PMB groups. The boronate **15** was prepared after a PMB protection of commercially available 4-bromobenzene-1,2-diol **14** followed by typical boronate formation in good yield.

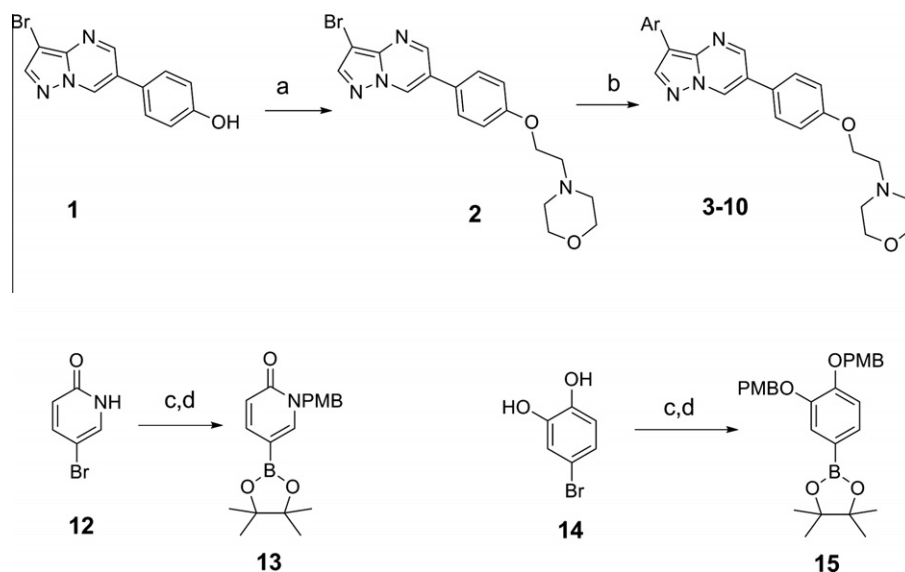
Two synthetic routes were developed for analog **5**. The overall reaction in Scheme 1 was followed. The boronate **13** was prepared after PMB protection of compound **12**, followed by standard boronate formation in good yield. The product was then reacted with bromide **2** under Suzuki cross coupling conditions to give the desired analog **5** after deprotection. Inhibitor **9** was also converted to analog **5** via a one pot two step synthesis consisting of diazotization and hydrolysis in moderate yield as outlined in Scheme 2.<sup>20</sup>

The inhibitory activities of all analogs were tested against the hAMPK  $\alpha 2$  subunit in complex with the heterotrimer (hAMPK  $\alpha 2$ ,  $\beta 1$ ,  $\gamma 1$ ). The structure–activity relationship of the pyrazolo[1,5-*a*]pyrimidines is summarized in Table 1. Synthetic intermediates **1** and **2** were included to test the contribution of each fragment on the binding of the pyrazolo[1,5-*a*]pyrimidines to AMPK. The addition of the morpholinoethyl resulted in a twofold improvement in potency observed in **2** relative to **1**. Replacement of the piperidine tail in compound C (Fig. 1) with the morpholinoethyl group in **4** resulted in a sevenfold loss in potency. Additionally, the ADME filters used to prioritize the designed compounds for synthesis predicted that compounds with the morpholino fragment would have better human intestinal absorption.<sup>17</sup> The enhanced potency of **4** relative to **3** was attributed to the ability of the nitrogen on the pyridine ring to form a hydrogen bond with the side chain of Lys-45. We analyzed the results of the fragment simulation data to identify fragments that could form an additional hydrogen bond with the side chain of Asp-157 on the DFG-loop while retaining contact with Lys-45. Compounds **5**, **6**, **7**, **8**, **9**, and **11** were designed using this approach.

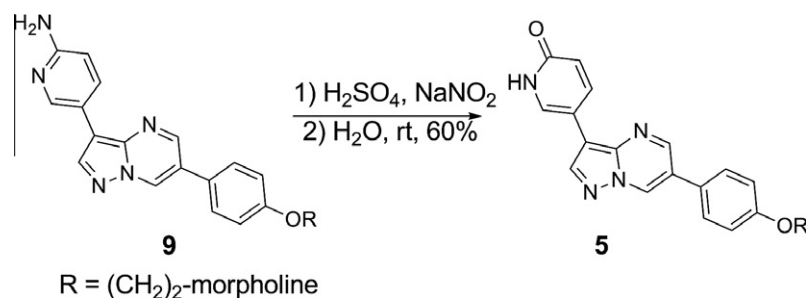
The pyridinone fragment of compounds **5** (Fig. 2) and **6** showed favorable interactions with the Lys-45/Asp-157 pair and two plausible attachment points to the pyrazolo[1,2-*a*]pyrimidine core.



**Figure 2.** Minimized predicted lowest-energy binding pose of compound C (A) and compound **5** (B) (carbons in magenta) in complex with the hAMPK  $\alpha 2$  subunit without their respective ethylpiperidinyl or ethylmorpholino tails. The hydrogen-bonds between the pyridinone oxygen and the zeta nitrogen of Lys-45; the pyridinone nitrogen and the delta oxygen of Asp-157 (B) and between the pyrazolopyrimidine core and the backbone nitrogen of Val-96 in the hinge region (A and B) are shown in green. Note that the DFG loop (orange) is in the 'in' conformation.



**Scheme 1.** Synthesis of pyrazolo[1,5-*a*]pyrimidine compounds. Reagents and conditions: (a)  $K_2CO_3$ , DMF, 80 °C, 90%, 4-(2-chloroethyl)morpholine HCl; (b) 10 mol %  $PdCl_2(PPh_3)_2$ ,  $Na_2CO_3$  2 M,  $ArB(OH)_2$ ; (c) PMBCL, cat. NaI, DCM, rt, 80%; (d) 10%  $PdCl_2$  dppf.dcm, Bis (pinacolato) diborane, NaOAc, DMF, 80 °C, 75%.



**Scheme 2.** Alternative synthetic route to **5**.

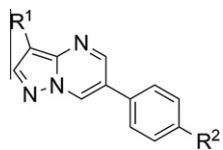
Both variants were made to test the preference for the geometry of the Lys-45/Asp-157 interaction. Compound **6** showed no improvement over **4**, while **5** displayed equipotent activity compared to compound C. Compound **7** displayed twofold improvement in activity relative to **4**. As with the pyridinone fragment, the GCMC simulations indicated that the 2-aminopyridine fragment would form profitable interactions with the Lys-45/Asp-157 pair, and two compounds could be designed with different attachment points to the pyridine ring that preserve the hydrogen bonds. Compound **8** was found to be equipotent with **4**, while **9** was 10-fold weaker. The catechol was also identified as a potentially favorable fragment that might preserve the key interactions, so **11** was synthesized but found to be fourfold weaker than **4**. The advanced intermediate **10**, lacking a hydrogen bond donor, was twofold less potent than the target compound **11**. Another reason for the reduced potency of **10** relative to **11** is likely due to steric clashes between the methoxy groups and the side chain atoms of either Lys-45 or Asp-157. Taken together, the resulting SAR for the pairs of **5** and **6** as well as **8** and **9** demonstrate a clear preference for the trajectory from the core for the hydrogen bond acceptor and donor atoms, while compounds **5** and **7** show that the addition of a hydrogen bond donor to exploit the interaction with Asp-157 can result in improved potency against hAMPK.

We investigated the replacement of the pyrazolo[1,5-*a*]pyrimidine core by searching for fragments that would form favorable interactions with the hinge region while preserving the orientation of fragments that were believed to form favorable contacts with Lys-45 and Asp-157. Several core replacements suggested by the

fragment simulations were used to design new compounds with reduced molecular weight that preserved the hydrogen bonding interaction with the backbone nitrogen of Val-96. These included 2-aminooxazoles (Table 2), pyridines, pyrimidines, 2-aminothiazoles, 2-aminooxazoles, pyrazolo[1,5-*a*]pyrimidine-7-amines, and pyrazolo[1,5-*a*]pyrimidin-2-amines. Due to synthetic feasibility concerns, only the designs containing pyridine, 2-aminooxazole, and pyrazolo[1,5-*a*]pyrimidine-7-amine cores were synthesized and tested. The pyridine and pyrazolopyrimidine-7-amine cores showed weak or no activity ( $>10 \mu M$ ) in the APMK assay (data not shown). The most active of these core replacements was the 2-aminooxazole series. The 2-aminooxazoles are predicted to bind to AMPK with the oxazole ring nitrogen forming the hydrogen bond with the backbone of Val-96. The 2-amino group is predicted to form an additional hydrogen bond contact with the backbone carbonyl of Ser-97. Replacements for the 2-(piperidin-1-yl)ethoxy fragment were also investigated and the most promising one identified from the GCMC simulations was benzene sulfonamide. The oxygen of the sulfonamide was predicted to form a hydrogen bond with the backbone nitrogen of Glu-100, while the nitrogen acts as a hydrogen bond donor to a side chain oxygen of the same residue.

The synthesis of the 2-aminooxazole compounds using an iminophosphorane-mediated cyclization is outlined in Scheme 3.<sup>21</sup> The reaction of an isothiocyanate with an acyl azide and triphenylphosphine in dioxane at 95 °C or in dichloromethane at room temperature provided the 2-aminooxazole.<sup>22</sup> The acyl azide **20** was obtained from the available 2-chloro-1-(3,4-dihydrophenyl)ethanone via a Finkelstein reaction with sodium azide.<sup>23,24</sup>

**Table 1**  
Inhibition of hAMPK ( $\alpha 2$ ,  $\beta 1$ ,  $\gamma 1$ ) by pyrazolopyrimidines



No.	R <sup>1</sup>	R <sup>2</sup>	IC <sub>50</sub> <sup>a</sup> (μM)
1	Br	OH	7.7
2	Br	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	3.5
3	4-Me-Ph	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	1.8
4	4-Me-Py	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	0.28
5	4-Me-2-Pyridone	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	0.06
6	4-Me-2-Pyridone	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	0.27
7	4-Me-2-Pyridone	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	0.14
8	4-Me-2-Pyridone	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	0.22
9	4-Me-2-Pyridone	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	2.1
10	4-Me-2-Pyridone	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	2.2
11	4-Me-2-Pyridone	CH <sub>2</sub> CH <sub>2</sub> N(CH <sub>2</sub> ) <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> OH	1.1

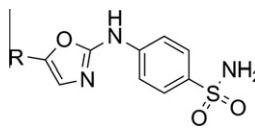
<sup>a</sup> Values were measured using the Hot Spot™ filtration binding <sup>33</sup>P Kinase assay, Reaction Biology, Malvern, PA. Compounds were tested in ten-dose IC<sub>50</sub> mode with threefold dilutions starting at 10 μM. All reactions were carried out in the presence of 10 μM ATP.

The acyl azide intermediate **19** was synthesized from the corresponding acyl bromide **18** as shown in Scheme 4. Compound **17** was generated via Stille cross-coupling from **16** (see also Scheme 1).<sup>25,26</sup> Compounds **19** and **20** were converted to the intermediate iminophosphorane upon treatment with PPh<sub>3</sub>, which was followed by rearrangement and deprotection to yield **21** and **22**.

Compound C has previously been reported to be a potent inhibitor of a number of kinases in addition to AMPK, including Eph-A2, c-Src, Lck, KDR, and MNK-1.<sup>15</sup> A kinase selectivity screen was carried out using these six kinases plus the relevant cancer kinases Flt1, Flt3, and Rsk1. We tested the most potent pyrazolo[1,5-*a*]pyrimidines in addition to two 2-aminooxazoles along with compound C. As seen in Table 3, compound C is non-selective against the kinases in screen, inhibiting seven of the eight kinases more potently than AMPK. Compound **5**, which is equipotent with compound C for AMPK, is substantially more selective. With the exception of Flt3, **5** showed improvements in selectivity between 4.5-fold for Mnk1 and 60-fold for Flt1 as compared to compound C.

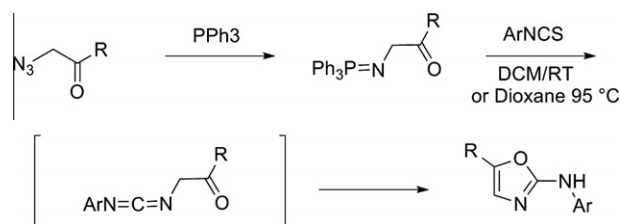
Figure 3 compares the effect of **5** and compound C on basal AMPK activity (detected by immunoblotting for specific phosphorylation of the AMPK substrate acetyl CoA carboxylase 1 (ACC) on

**Table 2**  
Inhibition of hAMPK  $\alpha 2$  by aminooxazoles

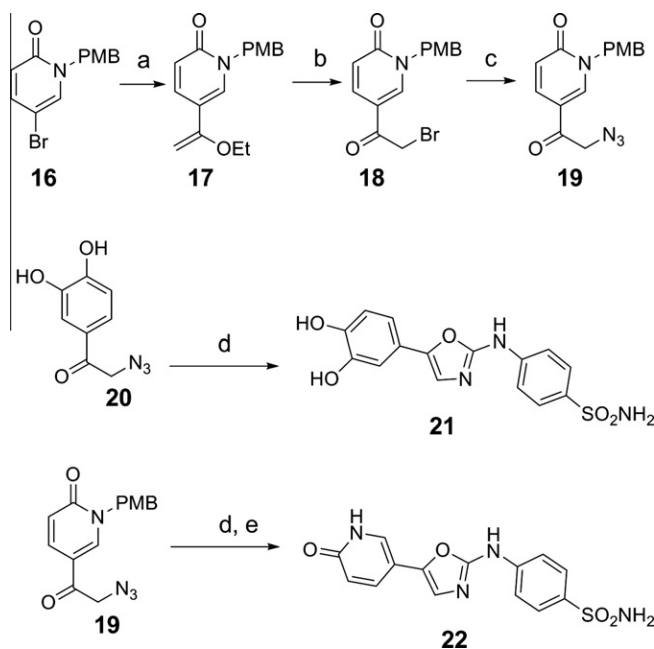


No.	R	IC <sub>50</sub> <sup>a</sup> (μM)
21	3,4-Di-OH-Ph	2.9
22	4-Me-2-Pyridone	5.0

<sup>a</sup> As in Table 1.



**Scheme 3.** General method for the synthesis of 2-aminooxazoles.



**Scheme 4.** Synthesis of 2-aminooxazoles. Reagents and conditions: (a) 10 mol % PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub>, LiCl, vinyltin reagent; (b) NBS THF/water; (c) NaN<sub>3</sub> acetone/water; (d) PPh<sub>3</sub>, ArNCS; (e) TFA, 100 °C, microwave.

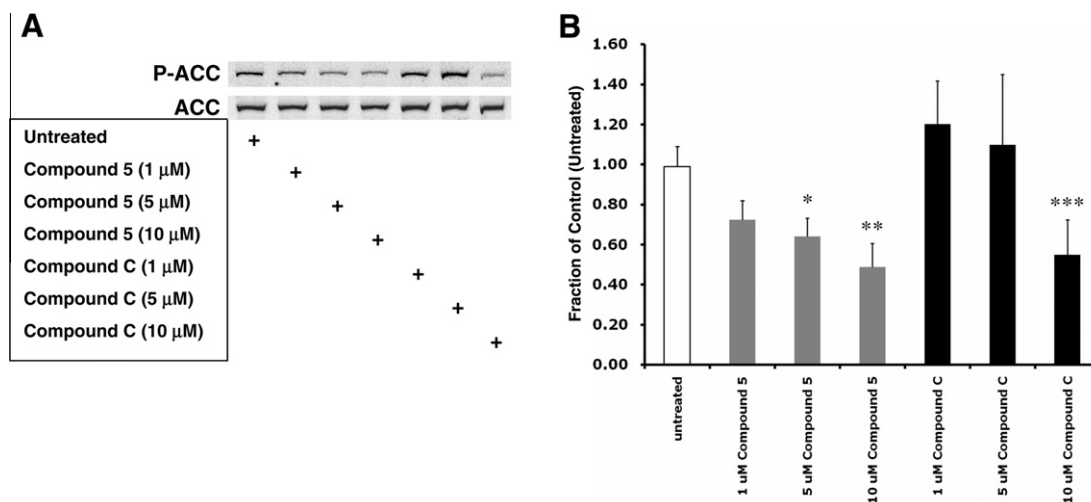
Ser79; Phospho-ACC<sup>3,27</sup>) in a mouse embryonic fibroblast (MEF) cell line. Transformed derivatives of these cells were previously used to investigate the contribution of AMPK to experimental tumor growth.<sup>3</sup> Both **5** and compound C inhibited cellular AMPK activity (P-ACC levels), but **5** was a more potent AMPK inhibitor than compound C (e.g., ≤5 μM, 6 h) in MEF cells. It is noteworthy that both **5** and compound C appeared to decrease specific phosphorylation of the AMPK $\alpha$  catalytic subunit (P-AMPK levels; results



**Table 3**

Near kinase selectivity panel of pyrazolopyrimidines and aminooxazoles.

No.	IC <sub>50</sub> <sup>a</sup> (μM)								
	hAMPKα2	c-Src	EphA2	Flt1	Flt3	KDR	Lck	Mnk1	Rsk1
Compound C	0.041	0.002	0.011	0.011	<0.001	0.004	0.016	0.011	0.21
<b>4</b>	0.27	0.050	0.25	0.21	0.001	0.056	0.30	0.13	2.1
<b>5</b>	0.061	0.061	0.26	0.47	<0.001	0.10	0.43	0.045	2.1
<b>7</b>	0.14	3.3	0.30	0.004	<0.001	0.010	0.094	0.027	0.22
<b>8</b>	0.22	0.021	0.84	0.040	0.001	0.058	0.44	0.164	1.1
<b>11</b>	1.1	0.096	0.74	0.002	0.001	0.001	0.02	0.95	0.56
<b>21</b>	2.8	3.2	>10	0.074	0.13	0.100	0.97	2.2	3.4
<b>22</b>	5.0	8.2	5.6	>10	0.12	>10	>10	1.10	>10

<sup>a</sup> As in Table 1.

**Figure 3.** Effects of compound **5** and compound C on AMPK-dependent ACC phosphorylation (P-ACC) and phosphorylation of AMPKα on Thr172 (P-AMPK) in MEFs cultured in complete medium (DMEM + 25 mM HEPES [pH 7.4] + 10% fetal bovine serum [FBS]). **A** Representative immunoblots of total protein from MEFs harvested following exposure to compound **5** or compound C (1, 5, or 10 μM; 6 h). Replicate blots were probed for the relative levels of P-ACC or total ACC. **B** Corresponding histogram of densitometry measurements from multiple immunoblots of total protein from untreated (control) MEFs and MEFs treated with compound **5** or compound C; data were normalized to the control data points. Error bars, ±SD from independent cultures/data point ( $n = 3$ ). Statistical comparisons were made between control and treated cells (2-tailed unpaired  $t$ -test;  $p \leq 0.05$  was considered a significant difference); only significant differences between control and treated cells are shown (\* $p = 0.03$ , 5 μM compound **5**; \*\* $p = 0.03$ , 10 μM compound **5**; \*\*\* $p = 0.04$ , 10 μM compound C). Immunoblotting protocols have been described in detail.<sup>28</sup>

not shown) in these cells; it is not clear, however, whether this effect is direct or indirect.<sup>28</sup> In summary, the results shown in Figure 3 demonstrate that **5** inhibited endogenous AMPK activity in cultured mammalian cells.

In conclusion, pyrazolo[1,5-*a*]pyrimidine and 2-aminooxazole inhibitors of the human AMPKα2 subunit were identified through the use of FBDD and medicinal chemistry efforts. A binding model was obtained from the results of GCMC fragment simulations in to a homology model of the protein and is consistent with the observed SAR. The most potent pyrazolo[1,5-*a*]pyrimidine is equipotent with compound C in the in vitro AMPK assay and shows improved kinase selectivity. Compound **5** also shows greater inhibition against AMPK activity in a cellular assay relative to compound C.

## Acknowledgements

This work was supported by the Grants CA132529 and CA73807 from the NIH, National Cancer Institute. We would like to thank Dr. Kevin Moriarty and Dr. Martha Kelly for helpful discussions and critical review of this Letter.

## References and notes

- Mishra, R.; Cool, B. L.; Laderoute, K. R.; Foretz, M.; Viollet, B.; Simonson, M. S. *J. Biol. Chem.* **2008**, *283*, 10461.
- Pang, T.; Zhang, Z.-S.; Gu, M.; Qiu, B.-Y.; Yu, L.-F.; Cao, P.-R.; Shao, W.; Su, M.-B.; Li, J.-Y.; Nan, F.-J.; Li, J. *J. Biol. Chem.* **2008**, *283*, 16051.
- Laderoute, K. R.; Amin, K.; Calaoagan, J. M.; Knapp, M.; Le, T.; Orduna, J.; Foretz, M.; Viollet, B. *Mol. Cell. Biol.* **2006**, *26*, 5336.
- Papandreou, I.; Lim, A. L.; Laderoute, K.; Denko, N. C. *Cell Death Differ* **2008**, *15*, 1572.
- Viollet, B.; Horman, S.; Leclerc, J.; Lantier, L.; Foretz, M.; Billaud, M.; Giri, S.; Andreelli, F. *Crit. Rev. Biochem. Mol. Biol.* **2010**, *45*, 276.
- Clark, M.; Meshkat, S.; Wiseman, J. S. *J. Chem. Inf. Model.* **2009**, *49*, 934.
- Clark, M.; Guarnieri, F.; Shkurko, I.; Wiseman, J. S. *J. Chem. Inf. Model.* **2006**, *46*, 231.
- Clark, M.; Meshkat, S.; Talbot, G.; Carnevali, P.; Wiseman, J. S. *J. Chem. Inf. Model.* **2009**, *49*, 1901.
- Guarnieri, F.; Mezei, M. *J. Am. Chem. Soc.* **1996**, *118*, 8493.
- Carnevali, P.; Toth, G.; Toubassi, G.; Meshkat, S. *N. J. Am. Chem. Soc.* **2003**, *125*, 14244.
- Zhou, G.; Myers, R.; Li, Y.; Chen, Y. Z.; Shen, X.; Fenyk-Melody, J.; Wu, M.; Ventre, J.; Doebber, T.; Fujii, N.; Musi, N.; Hirshman, M. F.; Goodyear, L. J.; Moller, D. E. *J. Clin. Invest.* **2001**, *108*, 1167.
- Chemical Computing Group Inc.: Montreal, Quebec, Canada, 2006.
- Littler, D. R.; Walker, J. R.; Davis, T.; Wybenga-Groot, L. E.; Finerty, P. J., Jr.; Newman, E.; Mackenzie, F.; Dhe-Paganon, S. *Acta Crystallogr., Sect. F* **2010**, *66*, 142.
- Marx, A.; Nugoor, C.; Muller, J.; Panneerselvam, S.; Timm, T.; Bilang, M.; Mylonas, E.; Svergun, D. I.; Mandelkow, E. M.; Mandelkow, E. *J. Biol. Chem.* **2006**, *281*, 27586.
- Bain, J.; Plater, L.; Elliot, M.; Shpiro, N.; Hastie, C. J.; McLauchlan, H.; Klevernic, I.; Arthur, J. S.; Alessi, D. R.; Cohen, P. *Biochem. J.* **2007**, *408*, 297.
- Moore, W. R., Jr. *Curr. Opin. Drug Disc. Dev.* **2005**, *8*, 355.
- Klon, A. E.; Lowrie, J. F.; Diller, D. J. *J. Chem. Inf. Model.* **2006**, *46*, 1945.
- Churcher, I.; Hunt, P. A.; Stanton, M. G. WO/2007/085873, 2007.
- Cuny, G. D.; Yu, P. B.; Laha, J. K.; Xing, X.; Liu, J.-F.; Lai, C. S.; Deng, D. Y.; Sachidanandan, C.; Bloch, K. D.; Peterson, R. T. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 4388.

20. Daab, J. C.; Bracher, F. *Monatsh. Chem.* **2003**, *134*, 573.
21. Dhar, T. G. M.; Shen, Z.; Guo, J.; Liu, C.; Watterson, S. H.; Gu, H. H.; Pitts, W. J.; Fleener, C. A.; Rouleau, K. A.; Sherbina, N. Z.; McIntyre, K. W.; Witmer, M. R.; Tredup, J. A.; Chen, B.-C.; Zhao, R.; Bednarz, M. S.; Cheney, D. L.; MacMaster, J. F.; Miller, L. M.; Berry, K. K.; Harper, T. W.; Barrish, J. C.; Hollenbaugh, D. L.; Iwanowicz, E. J. *J. Med. Chem.* **2002**, *45*, 2127.
22. Froeyen, P. *Phosphorus, Sulfur, Silicon Relat. Elem.* **1991**, *60*, 81.
23. Chung, F.; Tisné, C.; Lecourt, T.; Dardel, F.; Micouin, L. *Angew. Chem., Int. Ed.* **2007**, *46*, 4489.
24. Sutton, J. C.; Pi, Z.; Rejean, R.; L'Heureux, A.; Thibeault, C.; Lam, P. Y. S. U.S. Patent WO/2006/078621, 2006.
25. Mitchell, T. N. In *Metal-Catalyzed Cross-Coupling Reactions*; Armin de Meijere, P. D. F. D., Ed., 2nd ed.; Wiley, 2008; p 125.
26. Farina, V.; Roth, G. P. *Adv. Metal-Org. Chem.* **1996**, *1*, 1.
27. Hardie, D. G.; Pan, D. A. *Biochem. Soc. Trans.* **2002**, *30*, 1064.
28. Laderoute, K. R.; Calaoagan, J. M.; Madrid, P. B.; Klon, A. E.; Ehrlich, P. J. *Cancer Biol. Ther.* **2010**, *10*, 68.